New Preorganized γ-Amino Acids as Foldamer Building Blocks

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Peptidic oligomers that contain γ -amino acid residues have become increasingly popular as subjects for foldamer research.¹⁻³ γ-Amino acid building blocks in which a ring constrains at least one backbone bond are of particular interest because cyclic constraints have proven to be useful in stabilizing specific secondary structures among foldamers that contain β -amino acid residues.⁴ We previously found that a five-membered ring incorporated via C_α and C_{γ} promotes a *γ*-peptide sheet secondary structure,⁵ and comparable observations have been reported by Smith et al.

for γ residues in which a cyclopropyl ring links C_{α} and C_{β} ⁶ More recently we have shown that a γ residue of type I (Figure 1), with a cis cyclohexyl constraint at the $C_\beta - C_\gamma$ bond (θ torsion angle), supports helical secondary structures among γ -, α/γ -, β/γ -, and $\alpha/\beta/\gamma$ -peptides.⁷ Identification of additional $γ$ -containing foldamers will require the development of a methodology to prepare new

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⁽¹⁾ Recent reviews: (a) Guichard, G.; Huc, I. Chem. Commun. 2011, 47, 5933. (b) Hecht, S., Huc, I., Eds. Foldamers: Structure, Properties and Applications; Wiley-VCH: Weinheim, Germany, 2007. (c) Seebach, D.; Beck, A. K.; Bierbaum, D. J. Chem. Biodiversity 2004, 1, 1111.

⁽²⁾ Selected γ-peptide examples: (a) Hanessian, S.; Luo, X.; Schaum, R.; Michnick, S. J. Am. Chem. Soc. 1998, 120, 8569. (b) Hintermann, T.; Gademann, K.; Jaun, B.; Seebach, D. Helv. Chim. Acta 1998, 81, 983. (c) Seebach, D.; Brenner, M.; Rueping, M.; Schweizer, B.; Jaun, B. Chem. Commun. 2001, 207. (d) Baldauf, C.; Gunther, R.; Hofmann, H. J. Helv. Chim. Acta 2003, 86, 2573. (e) Farrera-Sinfreu, J.; Zaccaro, L.; Vidal, D.; Salvatella, X.; Giralt, E.; Pons, M.; Albericio, F.; Royo, M. A. J. Am. Chem. Soc. 2004, 126, 6048. (f) Claudon, P.; Violette, A.; Lamour, K.; Decossas, M.; Fournel, S.; Heurtault, B.; Godet, J.; Mely, Y.; Jamart-Gregoire, B.; Averlant-Petit, M.-C.; Briand, J.-P.; Duportail, G.; Monteil, H.; Guichard, G. Angew. Chem., Int. Ed. 2010, 49, 333.

⁽³⁾ Selected examples with γ residues in heterogeneous backbones: (a) Hagihara,M.; Anthony, N. J.; Stout, T. J.; Clardy, J.; Schreiber, S. L. J. Am. Chem. Soc. 1992, 114, 6568. (b) Karle, I. L.; Pramanik, A.; Banerjee, A.; Bhattacharjya, S.; Balaram, P. J. Am. Chem. Soc. 1997, 119, 9087. (c) Ananda, K.; Vasudev, P. G.; Sengupta, A.; Raja, K. M. P.; Shamala, N.; Balaram, P. J. Am. Chem. Soc. 2005, 127, 16668. (d) Qureshi, M. K. N.; Smith, M. Chem. Commun. 2006, 5006. (e) Baldauf, C.; Gunther, R.; Hofmann, H. J. J. Org. Chem. 2006, 71, 1200. (f) Sharma, G. V. M.; Jadhav, V. B.; Ramakrishna, K. V. S.; Narsimulu, K.; Subash, V.; Kunwar, A. C. *J. Am. Chem. Soc.* 2006, 128, 14657. (g) Baruah, P. K.; Sreedevi, N. K.; Gonnade, R.; Ravindranathan, S.; Damodaran, K.; Hofmann, H. J.; Sanjayan, G. J. J. Org. Chem. 2007, 72, 636. (h) Vasudev, P. G.; Ananda, K.; Chatterjee, S.; Aravinda, S.; Shamala, N.; Balaram, P. J. Am. Chem. Soc. 2007, 129, 4039. (i) Chatterjee, S.; Vasudev, P. G.; Raghothama, S.; Ramakrishnan, C.; Shamala, N.; Balaram, P. J. Am. Chem. Soc. 2009, 131, 5956. (j) Araghi, R. R.; Jäckel, C.; Cölfen, H.; Salwiczek, M.; Vokel, A.; Wagner, S. C.; Wieczorek, S.; Baldauf, C.; Koksch, B. ChemBio-Chem 2010, 11, 335. (k) Araghi, R. R.; Baldauf, C.; Gerling, U. I. M.; Cadicamo, C. D.; Koksch, B. Amino Acids 2011, 41, 733.

Figure 1. Structures of γ and α amino acid residues.

γ-amino acid building blocks with diverse conformational constraints.

Here we describe asymmetric synthetic routes to γ -amino acids with a cyclohexyl constraint at the $C_{\alpha}-C_{\beta}$ bond (ζ torsion angle). Michael addition of nitromethane to cyclohexene-1-carboxaldehyde, facilitated via organocatalysis, provides access to protected derivatives of the cis and trans γ-amino acid diastereomers, each in highly enantioenriched form.⁸ Ultimately, this route enables us to incorporate γ residues of types II and III into α /γ-peptides. The *cis* residue (II) is shown to support helical secondary structure.

Our γ-amino acid synthesis efforts were motivated by results from Hayashi et al.⁹ and Wang et al.;¹⁰ these groups simultaneously reported that pyrrolidine A catalyzes a highly enantioselective Michael addition of nitromethane to β-substituted propenal derivatives. Use of benzoic acid as a cocatalyst proved to be necessary for high yields, and simple alcohols were superior as reaction media to nonpolar or polar aprotic solvents. Most of the reported examples involved $β$ -aryl propenals as the Michael acceptors; propenal derivatives bearing bulkier β -alkyl substituents required longer reaction times.⁹ It was unclear from these precedents whether cyclohexene-1-carboxaldehyde

(9) Gotoh, H.; Ishikawa, H.; Hayashi, Y. Org. Lett. 2007, 9, 5307.

Scheme 1

would be an effective Michael acceptor, since the enal unit bears both β-alkyl and α-alkyl substituents, which could sterically hinder the desired reactivity.

Reaction of nitromethane and cyclohexene-1-carboxaldehyde (3:1 ratio) in the presence of 20 mol $\%$ A and 10 mol $\%$ benzoic acid provided a pair of isomeric γ-nitro aldehyde products in an ∼4.5:1 ratio, based on NMR analysis (Scheme 1). The total yield was 90%. These aldehydes were immediately reduced to the corresponding *trans* and *cis* γ -nitro alcohols 1t and 1c; HPLC analysis indicated that each diastereomer was generated in >95% ee. Reduction was employed to avoid epimerization adjacent to the aldehyde group. The catalytic mechanism underlying the Michael addition presumably involves formation of an iminium ion via condensation of the starting aldehyde and A.

Diastereomers 1t and 1c were difficult to separate on a preparative scale, so the mixture of these δ -nitro alcohols was treated with $H_2Cr_2O_7$ in acetone, which quantitatively generated the corresponding γ-nitro carboxylic acids 2t and 2c. These diastereomers could be separated by chromatography and crystallization (major isomer 2t is a white solid, while 2c is an oil). The absolute configuration of 2t was established as (R, R) via the crystal structure of L-phenylalanine/ γ -dipeptide derivative 4 (Figure 2). We observed that the minor γ-nitro aldehyde stereoisomer generated by the Michael addition could be converted to the major γ -nitro aldehyde isomer by treatment with DBU, which suggests that γ -nitro carboxylic acid 2c has the S configuration at the ring carbon bearing the carboxyl group. This deduction was confirmed by oligomer crystal structures discussed below.

The high levels of enantiopurity generated for both product diastereomers in the Michael addition allowed us to evaluate the resulting γ -amino acids as foldamer building blocks. Boc-protected γ -amino acids 3t and 3c were readily prepared from γ -nitro acids 2t and 2c, respectively. We prepared α/γ -peptides with 1:1 α/γ alternation, $5-8$ (Figure 2), because computational studies by Hofmann et al.^{3e} and basic principles of conformational analysis suggested that such systems might display discrete

^{(4) (}a) Raguse, T. L.; Lai, J. R.; Gellman, S. H. J. Am. Chem. Soc. 2003, 125, 5592. (b) Schmitt, M. A.; Choi, S. H.; Guzei, I. A.; Gellman, S. H. J. Am. Chem. Soc. 2005, 127, 13130. (c) Horne, W. S.; Gellman, S. H. Acc. Chem. Res. 2008, 41, 1399. (d) Horne, W. S.; Johnson, L. M.; Ketas, T. J.; Klasse, P. J.; Lu, M.; Moore, J. P.; Gellman, S. H. Proc. Natl. Acad. Sci. U.S.A. 2009, 106, 14751. (e) Price, J. L.; Hadley, E. B.; Steinkruger, J. D.; Gellman, S. H. Angew. Chem., Int. Ed. 2010, 49, 368. (5) Woll, M. G.; Lai, J. R.; Guzei, I. A.; Taylor, S. J. C.; Smith,

M. E. B.; Gellman, S. H. J. Am. Chem. Soc. 2001, 123, 11077. (6) Qureshi, M. K. N.; Smith, M. Chem. Commun. 2006, 5006.

^{(7) (}a) Guo, L.; Chi, Y.; Almeida, A. M.; Guzei, I. A.; Parker, B. K.; Gellman, S. H. J. Am. Chem. Soc. 2009, 131, 16018. (b) Guo, L.; Almeida, A. M.; Zhang, W.; Reidenbach, A. G.; Choi, S. H.; Guzei, I. A.; Gellman, S. H. *J. Am. Chem. Soc.* **2010**, 132, 7868. (c) Sawada, T.; Gellman, S. H. J. Am. Chem. Soc. 2011, 133, 7336. (d) Guo, L.; Zhang, W.; Reidenbach, A. G.; Giuliano, J. W.; Guzei, I. A.; Spencer, L. C.; Gellman, S. H. Angew. Chem., Int. Ed. 2011, 50, 5843. (e) For an altertnative synthesis of γ amino acids of type I, see: Nodes, W. J.; Nutt, D. R.; Chippindale, A. M.; Cobb, A. J. A. J. Am. Chem. Soc. 2009, 131, 16016.

⁽⁸⁾ For racemic synthesis of γ -amino acids corresponding to residues II and III, see: Johnson,M. R.; Gauuan, J. F.; Guo, C.; Guzzo, P. R.; Le, V.-D.; Shenoy, R. A.; Hamby, J.; Roark, H.; Stier, M.; Mangette, J. E. Synth. Commun. 2011, 41, 2769. For asymmetric synthesis of the cyclobutane analogue of 3c, see: André, V.; Vidal, A.; Ollivier, J.; Robin, S.; Aitken, D. J. Tetrahedron Lett. 2011, 52, 1253.

⁽¹⁰⁾ Zu, L.; Xie, H.; Li, H.; Wang, J.; Wang, W. Adv. Synth. Catal. 2007, 349, 2660.

Figure 2. Structures of α/γ peptides 4–8 (arrows indicate H-bonds in the crystal structures 5 and 6).

folding propensities. Residues derived from γ -amino acid 3c (type II; as in α/γ -peptides 5 and 6) are expected to display gauche⁺ torsion angles about the θ and ζ backbone bonds. The computational studies predict that a local g^+ , $g⁺$ torsion angle sequence should promote formation of an α/γ -peptide helix that contains C=O(i)--H-N(i+3) H-bonds. This "12-helix" (so-named because the H-bonds occur in 12-atom rings) has previously been documented among α/γ -peptides.^{3i,7a} In contrast, residues derived from γ-amino acid 3t (type III; as in α /γ-peptides 7 and 8) are expected to display an *anti* torsion angle for θ and a gauche⁻ torsion angle for ζ. The local a, g ⁻ torsion angle sequence, which is observed for the single γ residue of type III in dipeptide 4, is predicted to be consistent with an α/γ -peptide helix that contains C=O(i)--H-N(i-4) H-bonds.^{3e} This helix has never been documented experimentally. It should be noted that the absolute configuration is R at C_β of each γ residue in α/γ -peptides 4-7, because all of these residues derive from Michael reactions catalyzed by A (S configuration). On the other hand, L - α -amino acid residues were used for 5 and 6, but D - α amino acid residues were used for 7 and 8, based on the computational predictions of Hofmann et al.^{3e}

High-quality crystals of α/γ -peptides 5 and 6 were grown by slow evaporation of chloroform/heptanes/ether or chloroform/heptanes solutions, respectively, and the structures of these oligomers were solved based on X-ray diffraction data. Unfortunately, we were unable to crystallize either 7 or 8, despite extensive effort. Thus, at present we have no insight on the secondary structural propensities of these α/γ -peptides containing γ residues with the *trans* cyclohexyl constraint (type III).

The solid state conformations of α/γ -peptides 5 and 6 revealed two different helical secondary structures (Figure 3), both of which were first proposed by Hofmann et al. based on computational studies.^{3e} Tetramer 5 forms two intramolecular H-bonds. The H-bond involving the N-terminal Boc carbonyl conforms to the expected

Figure 3. Crystal structures of 5 (left), 6 (right): (top) views perpendicular to helical axes; (bottom) views along the helical axes.

 $C=O(i)$ --H-N($i+3$) pattern; however, the other H-bond has the pattern $C=O(i)$ --H-N(i-1), a 10-membered ring interaction. This secondary structure has been designated the 12/10-helix; the different orientations of the 12- and 10-membered ring H-bonds cause this helix to have only a small net dipole. The 12/10-helix has previously been proposed by Sharma, Kunwar et al. based on NMR analysis of α/γ -peptides containing acyclic γ residues.^{3f}

Hexamer 6 formed the expected 12-helix conformation in the solid state; all four possible $C=O(i)$ --H-N $(i+3)$ H-bonds were formed. The crystal contains three independent molecules, but all three conformations are very similar to one another. Balaram et al. have previously proposed a 12-helical conformation for α/γ -peptides in which the γ residues are derived from gabapentin (spiro cyclohexyl ring at C_β), based on NMR data.³ⁱ We have recently provided crystallographic evidence for the 12-helix in α/γ -peptides that contain γ residues with a *cis* cyclohexyl constraint on the $C_\beta - C_\gamma$ bond.^{7a}

Table 1 compares backbone torsion angles for the α and γ residues observed in crystal structures of α/γ -peptides 5 and 6, which contain *cis* γ residues (II), and the analogous data for the lone *trans* γ residue (III) in dimer 4. For comparison, backbone torsion angles are given for a previously reported 1:1 α/γ -peptide crystal structure that contains γ residues of type \mathbf{I}^{7a} ; this example contains the (R, R, R) -γ residue and D-alanine. Each of the γ residues in 6 displays a g^+, g^+ local conformation about the $C_\alpha - C_\beta(\zeta)$ and $C_{\beta}-C_{\gamma}(\theta)$ bonds, and ψ and ϕ torsion angles near -120° , with a somewhat wider distribution for the latter torsion angle. In contrast, the previously described γ residues of type I display g^-g local conformations. For both type I and type II γ residues, the crystal structures are consistent with the $C=O(i)$ --H-N($i+3$) H-bonded

Table 1. Backbone Torsion Angles (deg) for α/γ -Peptides

peptides	residues	ϕ	θ	ζ	ψ
1:1 α / γ -tetramer 5	α	-74.7			27.0
$(\gamma$ -residue II)	γ	64.5	47.5	45.0	-118.2
	α	-94.0			152.2
	γ	-98.0	59.7	60.2	-169.0
1:1 α /y-hexamer 6	α	-65.9			35.9
$(\gamma$ -residue II)	γ	-120.1	54.7	55.3	-129.7
	α	-70.3			-27.2
	γ	-138.3	58.6	56.2	-112.1
	α	-83.8			-28.5
	γ	110.5	167.9	-54.6	-64.4
(L) -Phe/ γ -dimer 4	α	-89.4			142.5
$(\gamma$ -residue III)	γ	105.5	-177.9	-54.1	108.6
1:1 α/γ-hexamer ^{7a}	α	75.5			22.8
(R, R, R) - γ residue I	γ	134.3	-57.7	-50.5	109.4
	α	56.3			41.1
	γ	134.7	-56.7	-55.7	109.8
	α	67.2			45.9
	γ	162.1	-55.4	-51.3	116.0
computational	α	72.3			28.8
study	γ	123.4	-52.6	-62.3	124.9
By Hofmann ^{3e}	α	69.8			29.1
	γ	123.3	-52.1	-62.7	122.6
	α	69.6			30.6
	γ	122.8	-53.8	-64.0	129.8

helical conformation for 1:1 α/γ -peptides predicted by Hofmann et al.^{3f} The α residue ψ torsion angles in our crystal structures (near -30° for L-Ala) are similar to those observed for a canonical 3_{10} -helix, which shares the $C=O(i)$ --H-N $(i+3)$ H-bonding pattern.

Average parameters for the $C=O(i)$ --H-N($i+3$) H-bonded helices formed by 1:1 α/γ -peptides were derived from the structural data as described previously¹¹ and are presented in Table 2. Each helical parameter was calculated from the α -carbons of four consecutive residues (for γ residues, C_β was used as an imaginary α -carbon). Nonhelical residues at C-termini were excluded from these calculations. Table 2 includes, for comparison, parameters

for two other helices that contain $C=O(i)$ --H-N($i+3$) hydrogen bonds: the 1:1 α/γ -peptide 12-helix containing γ residues of type I and the α-peptide 3₁₀-helix. The parameters for all three helices are quite similar, although it is intriguing that the 12-helix containing γ residues of type II has a somewhat smaller rise-per-turn than the 12-helix containing γ residues of type **I**.

We have developed a short, asymmetric route to γ amino acids that feature a cyclohexyl constraint on the $C_{\alpha}-C_{\beta}$ bond. Both *cis* and *trans* diastereomers can be produced in highly enantiopure forms. Preliminary evaluation of the foldamer potential of these γ-amino acids was carried out via synthesis and examination of oligomers with a 1:1 α/γ backbone. Crystal structures revealed a strong helix-forming propensity for the *cis* γ residue, but no insights could be gleaned regarding the *trans* γ residue. Further use of the *cis* γ residues for fundamental foldamer research would be facilitated by improvements in the synthetic route, since the required protected γ -amino acid is derived from the minor diastereomer produced in the key Michael addition. We anticipate that incorporation of the new γ residues into other types of heterogeneous peptidic backbones will give rise to new types of foldamers.

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Supporting Information Available. Full experimental details, characterization data for all compounds including ¹H and ¹³C NMR and crystallographic data for 5 and 6 (CIF). This material is available free of charge via the Internet at http://pubs.acs.org. The authors declare no competing financial interest.

^{(11) (}a) Barlow, D. J; Thorton, J. M. J. Mol. Biol. 1988, 5, 601. (b) Sugeta, H.; Miyazawa, T. Biopolymers 1967, 5, 673. (c) Kahn, P. C. Comput. Chem. 1989, 13, 185.

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